

## Experimental Evidence for Hydrogen Tunneling when the Isotopic Arrhenius Prefactor ( $A_H/A_D$ ) is Unity

Sudhir C. Sharma and Judith P. Klinman\*

Departments of Chemistry and Molecular and Cell Biology, University of California, Berkeley, California 94720-1460

Received August 11, 2008; E-mail: klinman@berkeley.edu

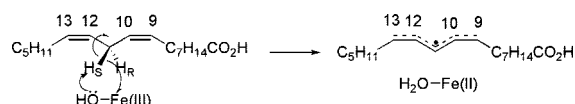
Elevated values for primary deuterium kinetic isotope effects ( $k_H/k_D$ ),<sup>1</sup> deviations of the temperature dependence of  $k_H/k_D$  from semiclassical predictions<sup>1</sup> and the breakdown of multiple isotope effects from simple reduced mass relationships (Swain–Schaad relationship)<sup>2,3</sup> have been used extensively in the detection of hydrogen tunneling near room temperature. In particular, the temperature dependence of  $k_H/k_D$  has emerged as an important diagnostic of H-tunneling, with isotopic Arrhenius prefactor ratios ( $A_H/A_D$ ) that are distinct from unity providing strong evidence for tunneling. However, the case is ambiguous when the value of  $A_H/A_D$  approaches unity (range of 0.8 to 1.4) and is generally interpreted as a reaction involving no tunneling.<sup>1</sup> Herein, we report experimental evidence for an isotopic Arrhenius prefactor ratio of unity in an enzymatic reaction that undergoes extensive tunneling of both protium and deuterium. Such an observation has been predicted within the environmentally coupled tunneling model put forth by Knapp et al.<sup>4</sup>

According to a Bell tunneling model,<sup>1</sup> all three hydrogen isotopes can cross a reaction barrier at some point below the classical transition state. Protium, with the longest de Broglie wavelength, can cross the barrier at a much lower and wider point than deuterium or tritium. A major consequence of this tunneling mechanism is an inflated deuterium kinetic isotope effect (KIE) that exceeds the semiclassical limit of  $\sim 7$ . At or near room temperature, where the majority of biological catalysts operate, this type of tunneling leads to the trend:  $E_a(\text{H}) < E_a(\text{D}) < E_a(\text{T})$ , and the corresponding trend in Arrhenius prefactors:  $A_H < A_D < A_T$ . There are a number of enzymatic reactions where the temperature dependence of experimental KIEs has been found to produce  $A_H/A_D$ ,  $A_H/A_T$  and/or  $A_D/A_T \ll 1$ .<sup>5</sup>

Arrhenius behavior deviations such that  $E_a(\text{H}) - E_a(\text{D}) \approx 0$ , leading to temperature-independent KIEs with  $A_H/A_D \gg 1$ , have also been observed for enzymes that transfer hydride, hydrogen atom, and proton.<sup>6</sup> These surprising, temperature-independent KIEs in enzyme-catalyzed reactions, together with the observation of Swain–Schaad deviations that cannot be accommodated within a Bell model,<sup>7</sup> have led to new theories for H-transfer in condensed phase that are much closer to Marcus models for electron transfer.<sup>4,8</sup>

The enzyme soybean lipoxygenase-1 (SLO-1) catalyzes the regio- and stereospecific conversion of linoleic acid (9,12-(Z,Z)-octadecanoic acid) (LA) to produce 13(S)-hydroperoxy-9(Z),11(E)-octadecadienoic acid [13-(S)HPOD]. The first part of this process is a proton-coupled electron transfer where the proton is transferred to the oxygen of the  $\text{Fe}^{3+}\text{-OH}$  and the electron is transferred to the iron center,<sup>9</sup> Scheme 1. Published kinetic data for wild-type (WT) SLO-1 indicate an extremely large KIE on  $k_{\text{cat}}$  ( $k_H/k_D = 81$ ) and a small Arrhenius prefactor and activation energy, together with experimental KIEs that are largely temperature-independent.<sup>10</sup> The crystal structure of SLO-1<sup>11</sup> indicates a substrate binding site lined by hydrophobic side chains, with the residues Leu<sup>546</sup> and Leu<sup>754</sup> in

**Scheme 1.** Mechanism of H• Abstraction Catalyzed by SLO-1



the vicinity of the active site cofactor  $\text{Fe}^{3+}\text{-OH}$ . A previous study demonstrated that single mutations, Leu<sup>546</sup>Ala and Leu<sup>754</sup>Ala, lead to a moderately temperature-dependent KIE and an elevated  $E_a$  in comparison to WT, whereas the more distal mutation, Ile<sup>553</sup>Ala, exhibits an enhanced temperature-dependent KIE with an inverse Arrhenius prefactor ratio.<sup>4</sup> A regular increase in the temperature dependence of the KIE following a progressive decrease in the bulkiness of the side chain at residue 553 was subsequently reported.<sup>12</sup>

The observation that the weakly temperature-dependent KIE of WT SLO-1 becomes more temperature-dependent upon the introduction of mutations at or near the active site has been explained within a Marcus-like, full tunneling model that allows for distance sampling (gating) between the H-donor and acceptor.<sup>4,8</sup>

$$k_{\text{tun}} = (\text{const}) \exp\left\{-\frac{(\Delta G^\circ + \lambda)^2}{4\lambda RT}\right\} \int_{r_1}^{r_0} \exp\left\{-\frac{m_H \omega_H r^2}{2\hbar J}\right\} \times \exp\{-E_x/k_b T\} dX \quad (1)$$

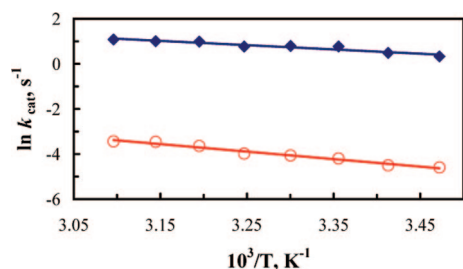
As represented in eq (1), the efficiency of hydrogen transfer is dependent upon three major factors: (1) A Marcus term that contains  $\lambda$ , the sum of outer and inner sphere reorganization to the reaction barrier, and  $\Delta G^\circ$ , the reaction driving force; this term is temperature dependent but only weakly isotope independent; (2) the Franck–Condon overlap, that defines the probability of wave function overlap between the H–donor and acceptor as a function of the mass ( $m$ ), frequency ( $\omega$ ), and distance traveled ( $r$ ) by the transferred hydrogen, and is strongly mass dependent; and (3) a donor/acceptor distance sampling term that reflects the barrier encountered,  $E_x$ , in reducing the distance between the H–donor and acceptor. The third exponential term connects both temperature and mass dependencies within a single function. Under this model, a close approach between the H–donor and acceptor distance, that ensures efficient wave function overlap for both protium and deuterium transfer, leads to similar enthalpies of activation for H• and D• transfer. However, when the donor/acceptor distance deviates from its optimal position, gating will impact the rate of D• transfer to a greater extent due to its smaller wavelength and subsequent poorer wave function overlap. This situation is especially apparent in the Ile<sup>553</sup>Gly mutant of SLO-1 where  $A_H/A_D = 0.027$  and the observed KIE is increased two-fold from that of the WT at 30 °C.<sup>12</sup>

In the case of WT SLO-1, the magnitude and nearly temperature-dependent KIEs implicate an optimized active site with a fixed and weakly modulated donor/acceptor distance that results from a relatively high gating frequency.<sup>4,12</sup> Introduction of active site

**Table 1.** Kinetic Parameters for SLO-1 and Mutants in 0.1 M Borate Buffer (pH 9.0)<sup>a</sup>

enzyme form	$k_{\text{cat}}^b$ (s <sup>-1</sup> )	KIE <sup>c</sup>	$E_a$ (H) (kcal/mol)	$\Delta E_a^d$ (kcal/mol)	$A_H$ (s <sup>-1</sup> )	$A_H/A_D$	$k_{\text{cat}}/K_M$ ( $\mu\text{M}^{-1}\text{s}^{-1}$ )
WT-SLO <sup>e</sup>	297 (12)	81 (5)	2.1 (0.2)	0.9 (0.2)	$9 \times 10^3$ ( $2 \times 10^3$ )	18 (5)	11 (1)
546A <sup>e</sup>	4.8 (0.6)	93 (9)	4.1 (0.4)	1.9 (0.6)	$4 \times 10^4$ ( $3 \times 10^4$ )	4 (4)	0.33 (0.1)
553A <sup>f</sup>	280 (10)	93 (4)	1.9 (0.2)	4.0 (0.3)	$7 \times 10^3$ ( $2 \times 10^3$ )	0.12 (0.06)	12 (1)
546A/553A <sup>g</sup>	2.21 (0.09)	128 (3)	3.8 (0.4)	2.8 (0.4)	$1.1 \times 10^3$ ( $5 \times 10^2$ )	1.05 (0.45)	0.11 (0.02)

<sup>a</sup> Data were collected from 15 to 50 °C. <sup>b</sup> The rate constants are reported for 30 °C. <sup>c</sup> KIE =  $k_{\text{cat}}^{\text{H}}/k_{\text{cat}}^{\text{D}}$ . <sup>d</sup> This is the isotope effect on  $E_a$ ,  $\Delta E_a = E_a(\text{D}) - \Delta E_a(\text{H})$ . <sup>e</sup> Reference 10a. <sup>f</sup> Reference 4a. <sup>g</sup> This work.



**Figure 1.** Arrhenius plot of kinetic data for Leu<sup>546</sup>Ala/Ile<sup>553</sup>Ala SLO-1 double mutant: data points for proto-linoleic acid [blue filled diamond (◆)] and dideutero linoleic acid [red open circle (○)]; linear fits to the Arrhenius equation are shown as solid lines; error bars are obscured by the symbol.

packing defects, via deletion of large hydrophobic side chains, alters the initial H–donor/acceptor distance as well as the oscillator frequency for distance modulation, causing enhanced temperature dependencies for the KIE.<sup>12,13</sup> Under this premise, two active site residues, Leu<sup>546</sup> and Ile<sup>553</sup>, have now been simultaneously mutated to Ala, and the corresponding hydrogen transfer parameters investigated.

Table 1 contains a summary of the kinetic data for the double mutant Leu<sup>546</sup>Ala/Ile<sup>553</sup>Ala, in relation to the respective single mutants Leu<sup>546</sup>Ala and Ile<sup>553</sup>Ala, as well as WT SLO-1. The overall rate of catalysis ( $k_{\text{cat}}$ ) of the double mutant Leu<sup>546</sup>Ala/Ile<sup>553</sup>Ala is decreased one hundred thirty-fold from that of WT and the single mutant Ile<sup>553</sup>Ala, and two-fold from the single mutant Leu<sup>546</sup>Ala. The rate of catalysis for the double mutant Leu<sup>546</sup>Ala/Ile<sup>553</sup>Ala between 15 and 50 °C is shown in Figure 1. The temperature dependencies of  $k_{\text{cat}}$  for both H• and D• abstraction were fitted to the empirical Arrhenius equation to yield  $E_a$  and the Arrhenius prefactor,  $A$ . This double mutant exhibits a more temperature-dependent isotope effect than WT or the single mutant Leu<sup>546</sup>Ala but less than the single mutant Ile<sup>553</sup>Ala. A particularly significant observation with this double mutant is that the magnitude of  $A_H/A_D$  is reduced to unity (1.05), an observation normally attributed to semiclassical hydrogen transfer. At the same time the experimental KIEs remain exceedingly large at all temperatures, at values much larger than permitted within a semiclassical H–transfer model. These observations show the difficulty of accommodating the experimental observations for the double mutant Leu<sup>546</sup>Ala/Ile<sup>553</sup>Ala using either semiclassical transition state theory or a Bell tunneling model.

By contrast, the environmentally coupled tunneling model, eq 1, requires at some point that  $A_H/A_D = 1$ , as a transition between a rigid optimized active site ( $E_a(\text{D}) - E_a(\text{H}) \approx 0$ ,  $A_H/A_D \gg 1$ ) and a site that has been compromised to the extent that extensive distance sampling becomes necessary to achieve a close enough distance between the H–donor and acceptor for efficient tunneling to occur ( $E_a(\text{D}) > E_a(\text{H})$ ,  $A_H/A_D \ll 1$ ). To our knowledge, the data presented herein provide the first unambiguous evidence for the

involvement of full quantum mechanical hydrogen tunneling for both protium and deuterium, where the isotopic Arrhenius prefactor ratio is unity. These results have important consequences for many reactions where the observation of  $A_H/A_D \approx 1$  has been automatically attributed to reaction via a semiclassical transition state. As an alternative, we posit that, within a given enzyme reaction, the trends of  $A_H/A_D$  provide a measure of the degree to which the H–donor/acceptor can achieve an optimal configuration for protium and deuterium wave function overlap. A range of  $A_H/A_D$  values may be expected that will fall between  $\gg 1$  and  $\ll 1$  depending on the impact of the environment on the tunneling efficiency. The behavior reported herein is expected to apply equally well to reactions characterized by relatively small isotope effects, as seen, for example in numerous enzyme-catalyzed hydride transfer reactions.<sup>14,6d,15–17</sup>

**Acknowledgment.** This work was funded in part by grants from the National Institutes of Health (Grant GM025765) and National Science Foundation (Grant MCB0446395).

**Supporting Information Available:** Experimental procedures: preparation of mutant enzyme, protein isolation and details of enzymatic assay. This information is available free of charge via the Internet at <http://pubs.acs.org>.

## References

- (1) Bell, R. P. *The Tunnel Effect in Chemistry*; Chapman and Hall: London; New York, 1980.
- (2) Swain, C. G.; Stivers, E. C.; Reuwer, J. F.; Schaad, L. J. *J. Am. Chem. Soc.* **1958**, *80*, 5885.
- (3) Cha, Y.; Murray, C. J.; Klinman, J. P. *Science* **1989**, *243*, 1325.
- (4) (a) Knapp, M. J.; Rickert, K.; Klinman, J. P. *J. Am. Chem. Soc.* **2002**, *124*, 3865. (b) Knapp, M. J.; Klinman, J. P. *Eur. J. Biochem.* **2002**, *269*, 3113.
- (5) (a) Grant, K. L.; Klinman, J. P. *Biochemistry* **1989**, *28*, 6597. (b) Whittaker, M. M.; Ballou, D. P.; Whittaker, J. W. *Biochemistry* **1998**, *37*, 8426. (c) Chowdhury, S.; Banerjee, R. *J. Am. Chem. Soc.* **2000**, *122*, 5417.
- (6) (a) Sikorski, R. S.; Wang, L.; Markham, K. A.; Rajagopalan, P. T. R.; Benkovic, S. J.; Kohen, A. *J. Am. Chem. Soc.* **2004**, *126*, 4778. (b) Francisco, W. A.; Knapp, M. J.; Blackburn, N. J.; Klinman, J. P. *J. Am. Chem. Soc.* **2002**, *124*, 8194. (c) Basran, J.; Sutcliffe, M. J.; Scrutton, N. S. *Biochemistry* **1999**, *38*, 3218. (d) Kohen, A.; Cannio, R.; Bartolucci, S.; Klinman, J. P. *Nature* **1999**, *399*, 496.
- (7) Klinman, J. P. *Phil. Trans. R. Soc.* **2006**, *B361*, 1323.
- (8) Kuznetsov, A. M.; Ulstrup, J. *Can. J. Chem.* **1999**, *77*, 1085.
- (9) Glickman, M. H.; Klinman, J. P. *Biochemistry* **1996**, *35*, 12882.
- (10) (a) Rickert, K. W.; Klinman, J. P. *Biochemistry* **1999**, *38*, 12218. (b) Jonsson, T.; Glickman, M. H.; Sun, S.; Klinman, J. P. *J. Am. Chem. Soc.* **1996**, *118*, 10319.
- (11) Tomchick, D. R.; Phan, P.; Cymborowski, M.; Minor, W.; Holman, T. R. *Biochemistry* **2001**, *40*, 7509.
- (12) Meyer, M. P.; Tomchick, D. R.; Klinman, J. P. *Proc. Natl. Acad. Sci. U.S.A.* **2008**, *105*, 1146.
- (13) The fact that the rate for H transfer is seen in selected mutants to vary very little in relation to WT, while D transfer is impeded, can be understood in the context of a compensating decrease in the force constant for gating that accompanies the increase in donor/acceptor distance. This aspect will be treated in greater detail elsewhere.
- (14) Wang, L.; Goodey, N. M.; Benkovic, S. J.; Kohen, A. *Proc. Natl. Acad. Sci. U.S.A.* **2006**, *103*, 15753.
- (15) Maglia, G.; Allemann, R. K. *J. Am. Chem. Soc.* **2003**, *125*, 13372.
- (16) Seymour, S. L.; Klinman, J. P. *Biochemistry* **2002**, *41*, 8747.
- (17) Tsai, S. C.; Klinman, J. P. *Biochemistry* **2001**, *40*, 2303.

JA806354W